

工學碩士 學位論文

高農度窒素 含有 重金屬廢水
生物學的處理

**Biological Treatment of High concentrated
Nitrogen Wastewater with Heavy Metals**

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ABSTRACT

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ABSTRACT

Heavy metal pollution problems have become one of the most important environmental issues in Korea. This is because the metal contaminants can cause serious toxic effects in biota in aquatic and soil environments. Metal plating wastewater has been usually treated by the physico-chemical methods such as chemical precipitation, reverse osmosis and evaporation. Characteristics of the wastewater, however, became increasingly complicated as the industry developed and combined treatment of the wastewater for the small scale factories became more common. Treatment of metal plating wastewater requires more effective technologies in terms of performance and economics that would be applied to the small scale metal industries

The long term goal of this study was to develop a metal plating wastewater treatment system that can be practically employed for the small scale metal industries. The specific aims of this study were : 1) to find and optimize the growth conditions of sulfate reducing bacterial consortia for an efficient removal, 2) to develop an anaerobic process packed with a floating granular media, 3) to develop a simultaneous removal process of heavy metals and nitrogen utilizing sulfate reducing bacteria(SRB), sulfur denitrification bacteria(SDNB), and heterotrophic denitrification bacteria(DNB).

To find effective treatment conditions for the electroplating wastewater treatment by SRB, the bacteria activity, removal capacity

of heavy metals, and possibility of nitrogen removal were investigated with respect to a wide range of COD/sulfate ratios in an anaerobic continuous reactor. During the start-up period when COD/sulfate ratios were gradually increased from 1.53 with the fixed COD concentration of 500 mg/L as glucose, successful sulfate reduction rate (above 95%) was achieved. Furthermore, in order to determine the activities of SRB at varying COD/sulfate ratios, influent COD concentrations were controlled to maintain COD/sulfate ratios at 0.18, 0.33, 0.5, 0.82, 1.2 after fixation of sulfate concentration of 2000 mg/L in the feed electroplating wastewater. At the relatively low values (0.18, 0.33) deficiency of organics in the feed affected the activity of SRB. Sulfate reduction efficiency was 61% and 59%, at the COD/sulfate ratio of 0.82 and 1.2, respectively. The further increase of the ratio did not appear to increase the reduction efficiency. Indicating sulfide inhibited the SRB's activity at the high COD/sulfate ratios. From the results, economic COD/sulfate ratio was determined to be 0.33 to reduce sulfate 2000 mg/L.

The amount of external carbon source supplied could be determined on the basis of heavy metal concentration in the wastewaters since sulfate reduction rate can be controlled by the external carbon. Heavy metals were effectively removed by SRB at above pH 6.4 regardless of metal species, but removal efficiency decreased dramatically at pH 5.4 to decrease of SRB activity caused by unfavorable pH condition. Consequently, at least, pH 5.4 or higher was necessary for the neutralization of electroplating wastewaters to maintain the stable activity of SRB.

In case of the heavy metal inhibition test in a nitrification reactor, the results of Phase 1 showed about 20% inhibition, and maximum inhibition up to 75% was observed at the later phase experiments. The removal possibility of ammonium nitrogen(400mg/L) in electroplating wastewaters was tested by utilizing a putative symbiotic relationship of three different microorganisms. Nitrates of high concentration effectively removed in a single anaerobic reactor containing alkalinity of 3000mg/L(as CaCO₃) and COD of 1000 (mg/L) by the putative symbiotic relationship of SRB, SDNB, and DNB.

The results from this study will contribute to an understanding of biological removal mechanism of heavy metals and nitrogen in the electroplating wastewater and to development of this treatment technologies in the future.

.

(SRB:Sulfate Reducing Bacteria)

(Propionate), (Butyrate), (Lactate) (Acetate),
VFA(Volatile fatty acids)

(MPB : Methan producing bacteria) 가

(19, 38).

(Sulfide) (Cr), (Cu),
(Metal Sulfide)
(Zn), (Pb) 가

(SCN) 가
(18,37,40,47). 가

(20),

가

i)

, ii)

iii)

, iv)

(12,21).

pH

(41,44,48).

가
가 가

(45).

2000mg/

가 가

가
(15,18,19,20).

(Heterotrophic Denitrifier)

가

(16).

가

(Autotrophic Denitrifier)

가가

가

(4,7,31). 가

가

가

가

.

, 가

,

가

가

.

(1).

가

가

가

가

(1).

가

가

,

가

가

가

가

.

•

2.1

가

. 가 ,

1

, , ,

27

,

24

(41,46).

가

500m³/d

4, 5

가

.

, ,

가

.

2.1.1

,

.

.

,

,

3

,

6 : 1.5 : 1.5

(45). Table 2.1

J

pH

Table 2.1 Characteristics of some wastewater sample from
J'electroplating union in Pusan(45).

Content	()			
COD	233.3	248.5	383.8	258
SS	7,438	10,942	7,668	8,030
TDS	6,876	10,838	7,050	7,496
pH	2.7	2.4	8.5	2.7
Sulfate	2,660	1,523	1,450	2,307
Chloride	5,400	9,000	5,000	5,880
TKN	325	755	350	393.25
CN	4.3	6.2	277.0	45.4
Cr+6	28.5	-	-	19.95
T - Cr	80.5	413.0	31.4	123.01
Cu	122.9	56.2	227.3	128.5
Fe	674.3	1027.0	252.1	663.87
Ni	128.8	82.2	86.0	115.39
Zn	380.6	647.5	219.0	396.39
Pb	3.9	7.7	4.6	4.57

가
가

1997

(45).

2.1.2

Fig. 2.1

가

, 100cm²

5 10

M₀

10m³/hr

(44,45,46).

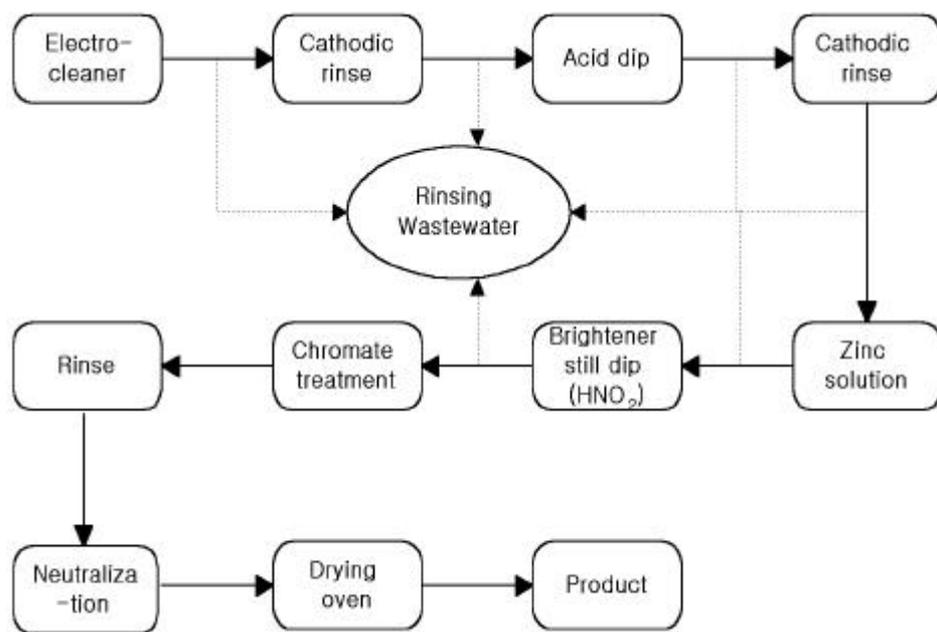


Fig. 2.1 Procedures of zinc plating(41,45).

2.2 (Sulfate Reducing Bacteria)

가

가 ,

가

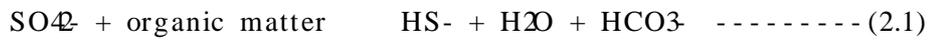
가 .

가

(18,19,37,40,43).

2.2.1 (Ecological Characteristics)

SRB



가

(19,37).

1) (Carbon Source)

H₂, Formate, Lactate, Acetate, Propionate, Butyrate, Higher Straight-chain Fatty Acids, Sugars

가

가

Table 2.2

(19,43).

Table 2.2 Stoichiometry of sulfate reducing reaction and methane producing reaction where hydrogen and acetate are used as electron donors(43).

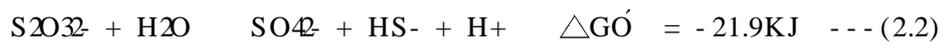
Reaction of methan producion	Reaction of sulfate reduing
$\text{CO}_2 + 4\text{H}_2 \rightarrow \text{CH}_4 + 2\text{H}_2\text{O}$ $G^0 = -135 \text{ kJ}$	$\text{SO}_4^{2-} + 4\text{H}_2 \rightarrow \text{H}_2\text{S} + 2\text{H}_2\text{O} + 2\text{OH}^-$ $G^0 = -154 \text{ kJ}$
$\text{CH}_3\text{COOH} \rightarrow \text{CH}_4 + \text{CO}_2$ $G^0 = -28.5 \text{ kJ}$	$\text{SO}_4^{2-} + \text{CH}_3\text{COOH} \rightarrow \text{H}_2\text{S} + 2\text{HCO}_3^-$ $G^0 = -43 \text{ kJ}$

2) (Sulfur Source)

(sulfite)

(thiosulfate)

(19).



Nielson(1991)

(2.2) ,(2.3)

가

가

(19).

3) (Dissolved Oxygen)

가

0.1- 1.0mg/

가 1.0mg/

(ORP) 가

가

가 (19).

4) (ORP)

가 - 100mV

가

가

가 - 100mV (19).

5) (Temperature)

10℃ 가

2 - 3.9 (

Q10. 9- 20℃

Q10 3.4 .

28- 32°C

45°C

(19).

6) pH

pH 5.5 pH 9 pH 7
7.5- 8 가 . SRB pH
(19).

7) (Sulfide)

가 . 가
1000mg/ 가
SRB
(Ferredoxin, Cytochrome)
(19).

2.2.2

가 가
. Hilton Oleszkiewicz(1998) H2S 가 200mg/
(19). Yoda(1987)
(19).
COD/SO₄ 가

COD/SO₄ 가 1.7

2.7

COD/SO₄ 가 1.7

2.7

0.4

(6,19). MacCartney

Oleszkiewicz(1993)

COD/SO₄ 가 3.7

(Propionate)

가 1.6

COD/SO₄ 가

가

(19,43).

2.2.3

(Metal interaction)

가

Table 2.4

가

Haas Polprasert(1994)

COD

SRB

가 가

Cr() Cr() SRB가

가

(15,18,19,37).

Mueller Steiner(1992)

Ni>Cu>Cd >Cr>Pb

(
 , pH,) .(19)

가 가 Cr()가 Cr()

가 (15,19).

SRB SEM AVS

. (, SEM: Simultaneously Extracted Metal, AVS:

Acid Volatile Sulfide) Cd SEM/AVS 가 1

가

CdS가 SEM/AVS

가 (19).

Table 2.3 Solubility products for various metal sulfides

and hydroxides(19).

Metal	K _{sp}	
	Metal sulfide	Metal hydroxide
Cu	8.5×10^{-45}	1.6×10^{-19}
Zn	1.2×10^{-23}	4.5×10^{-17}
Pb	3.4×10^{-28}	1.2×10^{-15}
Cd	3.6×10^{-29}	5.9×10^{-15}
Fe	3.7×10^{-19}	1.8×10^{-15}
Ni	1.6×10^{-16}	1.6×10^{-16}
Cr(III)	Not available	6.7×10^{-31}

2.3

2.3.1

. 1964
, 60 70 ,
.
가
(13,23,25).
, , ,
. 가
,
. 가
(heterotrophic metabolism) 가
- (13,23,25,34).
12 14% .
. 60% 40%
, 1%
NO3-N (25,34).

2.4 (Nitrification)

2.4.1 (Reaction by nitrification)

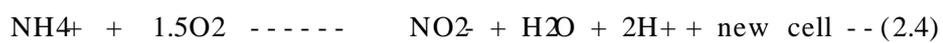
가 (Chemoautotrophic bacteria) .
 가 .
 가
Nitrosomonas . 가
Nitrobacter
 (13,25,34). Painter(1970) *Nitrospira*,
Nitrosococcus, *Nitrosolubus*, *Nitrosoglea*
 , *Nitrocystis*
Nitrosomonas
Nitrobacter .
 (28).

1) NH_4^+ NO_2^-

Nitrosomonas(, *N. europaea*)

hydroxylamine(NH_2OH)

(Nitrite) (13,23,25,34).

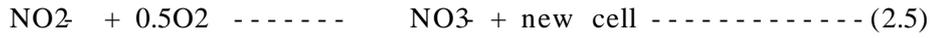


$$G(W) = 66 \text{ -- } 84 \text{ kJ/mol NH}_4^+\text{-N}$$

2) NO_2^- NO_3^-

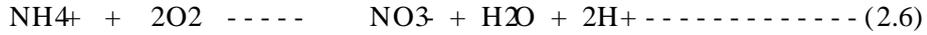
Nitrobacter(: *N. agilis*) (nitrate)

(13,23,25,34).



$$G(W) = 15.4 \quad 20.9 \text{ kJ/mol NO}_2\text{-N}$$

3)

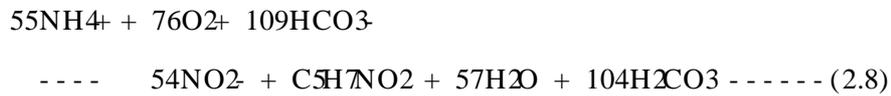


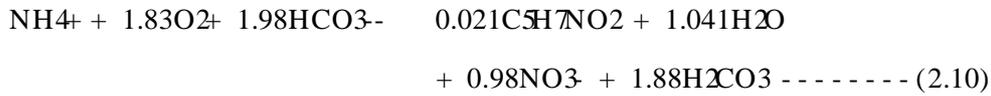
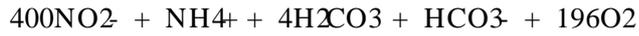
가 *Nitrosomonas* 66-84
 kcal/mole NH_4^+ *Nitrobacter*가
 17.5 kcal/mole NO_2^- .
 CO_2 , HCO_3^- , CO_3^{2-}

가
 가 .
 3.43g $\text{O}_2/\text{g N}$ 1.14g $\text{O}_2/\text{g N}$ 4.57g
 $\text{O}_2/\text{g N}$ (13,25,34).



(Yield Coefficient) -
 . *Nitrosomonas* *Nitrobacter*





Nitrosomonas Nitrobacter

0.15g VSS/g NH₄⁺-N 0.02g VSS/g NO₂⁻-N Alkalinity
 7.14g(as CaCO₃)/g NH₄⁺-N .

Nitrosomonas Nitrobacter 0.13g VSS/g NH₄⁺-N

0.02 0.07g VSS/g NO₂⁻-N

가

(maintenance)

Table

2.4 (13,23,25,34).

Table 2.4 Oxygen utilization, biomass yield, and alkalinity destruction coefficients acceptable for design of nitrification system[13,23,25].

	Coefficient
Oxygen demand (g O ₂ /g NH ₄ ⁺ -N)	4.6
Biomass yield (g VSS/g NH ₄ ⁺ -N)	0.1
Alkalinity reduce (g(as CaCO ₃)/g NH ₄ ⁺ -N)	7.1

Table 2.4 가 1g
 NO₃-N가 7.1 g 가 .
 가 NH₄⁺-N 가
 pH
 가 .

2.4.2

1) (Temperature dependency)

(maximum specific growth rate)

4 45 ,
Nitrosomonas 35 , *Nitrobacter* 35 42 . U.S.
 EPA(1975) *Nitrosomonas* (maximum growth rate)
 10 30

Table 2.4 . 30 35 Fig. 2.2
 (protein)

, 50

(25,28,29).

Table 2.5 Maximum specific growth rates for *Nitrosomonas sp.*

depending on temperature

Source	$\mu_{n,max}$ vs Temp.	$\mu_{n, max} (d^{-1})$		
		10	15	20
Dowing(1964a)	$(0.47)e^{0.08(T-15)}$	0.29	0.47	0.77
Dowing(1964b)	$(0.18)e^{0.116(T-15)}$	0.10	0.18	0.32
Hultman(1971)	$(0.50)10^{0.033(T-20)}$	0.23	0.34	0.50
Barnard(1975)	$0.33(1.127)e^{0.08(T-15)}$	0.10	0.18	0.37
Painter(1983)	$(0.18)e^{0.072(T-15)}$	0.12	0.18	0.26
Baccari(1979)				0.27
Bidstrup(1988)				0.65
Hall(1980)				0.46
Lawrence(1976)				0.50

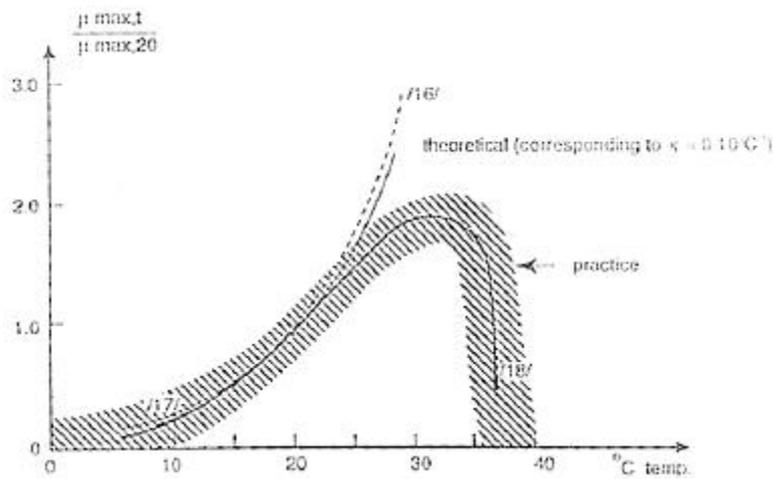


Fig 2.2 Nitrification as a function of temperature(25).

2) (Dissolved Oxygen concentration)

가 (Heterotrophic microorganism)
 . Dowing
 Scragg(1958) 0.3mg/L 가
 (10), Engel(1964) 가
 1mg/L *Nitrosomonas*
 2mg/L *Nitrobacter*
 (10). , Wuhrman DO
 가 4 7mg/L 1mg/L
 . DO
Nitrosomonas
 가 1mg/L
 , 2mg/L

3) pH

pH 가
 , pH . Fig.
 2.3 pH 7.5 pH .
 pH pH 7
 pH 8 pH 9 pH
 가 가 가 (25,29).

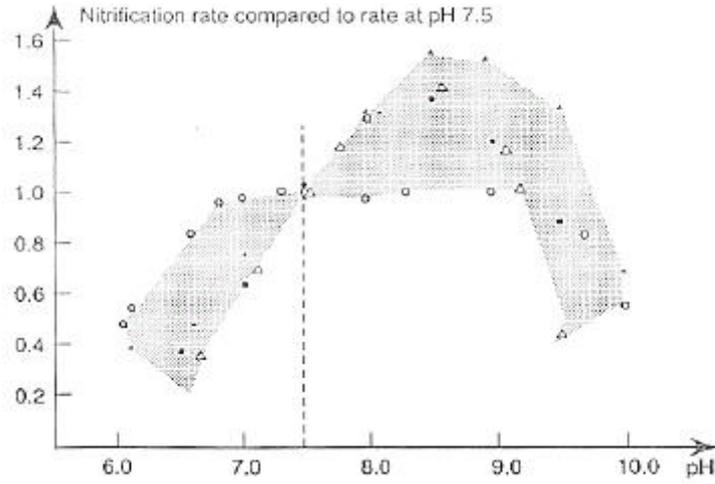


Fig. 2.3 Influence of pH on nitrification(25).

4)

가

Table 2.6

Free ammonia(FA $\text{NH}_3\text{-N}$), Free nitrous acid(FNA, HNO_2)

가

. FA *Nitrosomonas* 10 15mg/ , *Nitrobacter*
 0.1 1mg/L . FNA 0.22 2.8mg/
Nitrobacter Nitrosomonas . FA FNA
 NH_3 NH_4^+ $\text{NO}_2\text{-N}$, pH, ,
 가 (13,25,34).



Table 2.6 Organic compounds inhibiting nitrification(25).

Composition	Inhibition concentration(mg/L)	
	75%	50%
Acetone	2,000	-
Carbon disulfide	38	-
Chloroform	18	-
Ethanol	2,400	-
Phenol	5.6	-
Ethylenediamine	-	15
Hexamethylene diamine	-	85
Aniline	-	< 1
Monoethanolamine	-	> 200

pH 7 20 inhibitor
 NH₃, NH₄⁺ NO₂-N Threshold
 Table 2.7 . Table 2.7 가 가
 Nitrosomonas
 20% [13].
 Table 2.8 가

Table 2.7 Calculated threshold of ammonia plus ammonium nitrogen and nitrite plus nitrous acid nitrogen where nitrification inhibition may begin(pH 7, 20).

Inhibition conc(mg/L) for nitrification	Equilibrium conc. for NH ₃ -N + NH ₄ ⁺ -N	Equilibrium conc. for NO ₃ ⁻ -N + NO ₂ ⁻ -N
FA(Free Ammonia)		
10 for Nitrosomonas	1000	-
0.1 for Nitrobacter	20	-
FNA(Free Nitrous Acid)		
0.22 for Nitrification	-	280

Table 2.8 Concentration of nitrification inhibition with metals(25).

Metal	Concentration (g/m ³)	Effect
Cu	0.05- 0.56	Nitrosomonas activity inhibited(pure culture)
Cu	4	No essential inhibition in activated sludge
Cu	150	75% inhibition of activated sludge
Ni	>0.25	Nitrosomonas growth inhibited(pure culture)
Cr ³⁺	>0.25	Nitrosomonas growth inhibited(pure culture)
Cr ³⁺	118	75% inhibition of activated sludge
Zn	0.08-0.5	Inhibition of Nitrosomonas(Pure culture)
Co	0.08-0.5	Inhibition of Nitrosomonas(Pure culture)

2.5 (Denitrification)

가 . (anoxic)

가 가

N₂ Nitrous

oxide Nitric oxide 가

가

가

가

가

가

(facultive)

(terminal electron acceptor)

(13,25).

2.5.1 (Hetero denitrification)

(heterotrophic denitrifier) .

Gibbs > , >

가 , (13,25).

가 ,

1) DO (anaerobic denitrification) 가

(anoxic denitrification) 가

가 (13,25). Table 2.9

2 , 가 nitrogen gas

(Dissimilation) .

Table 2.9 Nitrogen Oxidation steps occurring for microbiological nitrogen removals.

Oxidation step for nitrogen	+5	+4	+3	+2	+1	0	-1	-2	-3
Assimilation	NO ₃ ⁻	-->	NO ₂ ⁻	-->	[NOH] ?	-->	NO ₂ O ₂ H	-->	R-NO ₂
Denitrification	NO ₃ ⁻	-->	NO ₂ ⁻	-->	NO ⁻	-->	N ₂		
Nitrification	NO ₃ ⁻	<--	NO ₂ ⁻	<--	[NOH]	<--	NO ₂ O ₂ H	<--	NH ₄ ⁺

(O2)

NO2--N NO3--N

가

pH

가

가 가

, (),

가

가

(25,34).

가

가

가

(Methanol),

(Ethanol),

(Acetic acid)

Table 2.10

. Fig 2.

가

(2.12) (13,23,25,34).



Table 2.10 Denitrification rates depending on the various carbon sources.

Various of carbon source	Denitrification rate (g NO3-N/g VSS-day)		Temperature()
	Methanol	0.21	
Sewage	0.12	0.90	20
	0.03	0.11	15 27
Endogenous metabolites	0.072	0.72	-
	0.017	0.048	12 20
Other carbon sources	Aceton, Molasses, Newsprint, Methane, Hydrogen(portable water), Wastewater from breweries, Olive oil, Acetic acid, Org. matter		

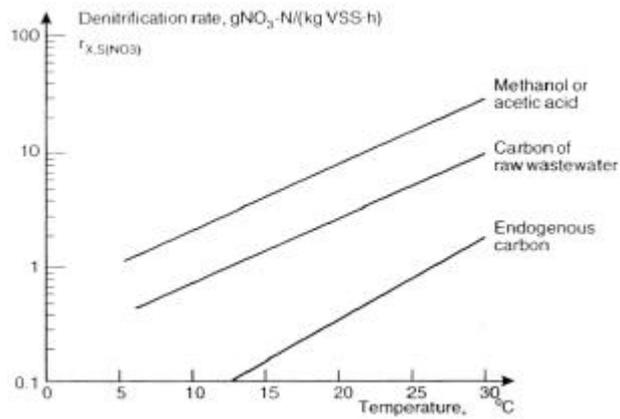


Fig 2.4 Influence of temperature on denitrification rate depending on different carbon sources(25).

(2.13)



1g

2.47g(COD

3.7g)

0.45g

3.57g

(13,25).

2.5.2

, DO,

, pH,

1) pH

가

가

가

pH가

pH 7.0 8.5 가
7.0 , pH가 8.0 9.5 7.0 4.0

1mg 가 3.57mg CaCO₃
가 3.0
pH
(13,25).

2) **(Dissolved oxygen)**

Skerman and MacRae(1957) *Pseudomonas*
DO 가 0.2mg/L DO가
[46], Nelson and Knowles(1978) DO가 0.13 mg/L
(26).

, 가
0.2mg/L 가 0mg/L
50% 가 .

3)

Monod .

4)

가
35 50 (5 1
0) . (50- 60) 35
50% 가 가

(13,25).

$$R_{DN(T)} = R_{DN(20)} K(T-20) \dots\dots\dots (2.14)$$

K 1.03 1.1 가 . 1.09

2.5.3 (Autotrophic denitrification)

Thiobasillus denitrificans *Thiomicrospira denitrificans*

(Sulfur, S-2) 가

(S₂O₃-2, S₄O₆-2, SO₃-2)

가

[3,4,7].

가

Beijrink

1904

가

(3,4,7).

C/N 가

가

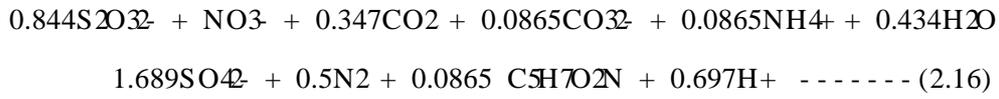
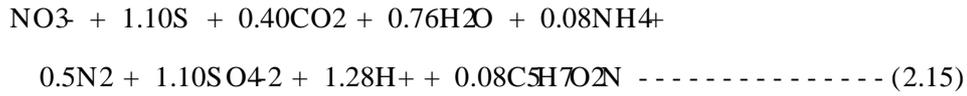
1)

(Autotrophic denitifier)

(2.15)

(2.16)

(4).



Sulfate . 2- 15 [SO₄²⁻/S-],
 [S-/NO₃⁻], [SO₄²⁻/NO₃⁻] 1, 1.1, 1.1 , (1.58) [SO₄²⁻/S₂O₃²⁻],
 [S₂O₃²⁻/NO₃⁻], [SO₄²⁻/NO₃⁻] 2, 0.844, 1.689 .

Table 2.11 Inhibition effects of ionic metals on
 denitrification efficiency(mg/L)(7).

Conc.(mg/L) Denitrification (%)	NO ₃ - N	NO ₂ - N	SO ₄ ²⁻	S ₂ O ₃ ²⁻	Na ⁺
100	2500	30	5000	11000	8300
0	6800	610	20000	50000	20700

2)

Claus Kutzner

가 Table 2.11

(7). 30 , pH 7.5 8.0

, NO₃- N S₂O₃²⁻ 10 20g/L

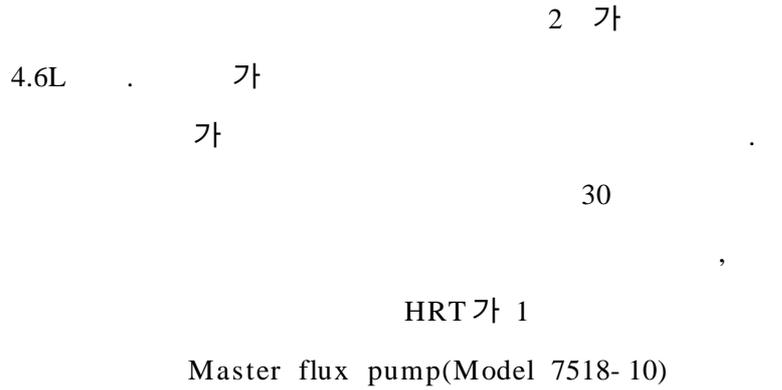
. 5g/L
 20g/L
 NO₂-N 가
 . Na+ 20g/L .
 pH 6 8 , pH
 , 가 ,
 Akalinity/NO₃-N 4
 Akalinity/NO₃-N 3
 (11).
 200mg/L .
 C/N 가 , 가
 가
 가 .

3.1

3.1.1

(Upflow Anaerobic Floating Media Biofilm Reactor)

Fig. 3.1



3.1.2

S 2L
Table 3.1

Table 3.1 Physical properties of porous ceramic supports

Size (mm)	Average pore diameter (μm)	Range of pore diameter (μm)	Porosity (%)	Bulk density (g/cm^3)
7 8	14 19	1.0 100	75 80	0.33 0.37

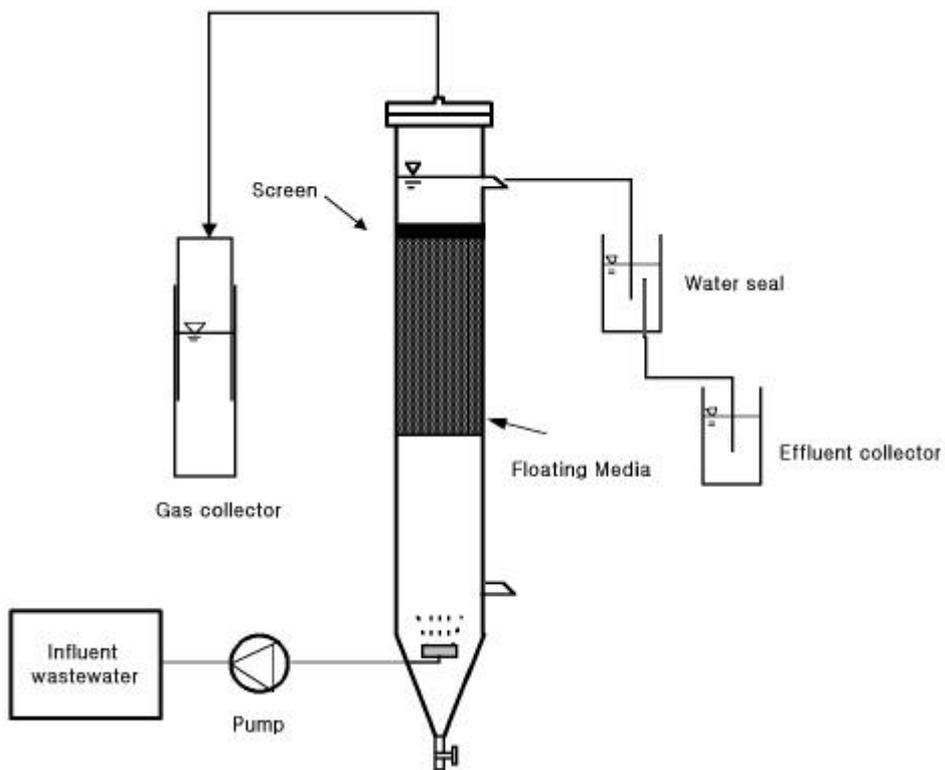


Fig. 3.1 Schematic diagram of upflow anaerobic floating media biofilm reactor.

3.1.3

50:50

VSS 11,500mg/L

2.6L

3.1.4

Fig3.2

2.6L

2

J

2L

Master flux pump(Model 7518- 10)

3.2

3.2.1

COD/Sulfate

1)

가

가

Table 3.2. Characteristics of the feeding wastewater in reactor A.

		COD	SO ₄ 2	NH ₄ N	PO ₄ P	Trace element
A	Start-up period	500	325- 500	400	80	Added**
	Steady state period	436- 1180	2000	400	80	"

Added** : Field et al.,[14].

2)

A COD/sulfate
 가 COD
 500mg/L
 가
 COD/sulfate
 (MgSO₄) J
 2000mg/L COD/sulfate가 0.18,
 0.33, 0.5, 0.82, 1.21가 COD 가

3.2.2

1)

가 pH
 가
 pH pH

가 pH

2) B pH 가

B

J Table 2.3

, pH (Ca(OH)₂ 54, 6.4, 7.2

B COD 1000mg/L

J NH₄Cl

400mg/L 가

pH

Table 3.3 Characteristics of the feeding wastewater in reactor B.

	COD	SO ₄ 2	NH ₄ N	PO ₄ P	Heavymetals Conc.					Trace element
					Cu	Fe	Ni	Zn	Cr	
B*	1000	2000	400	80	130	665	115	400	125	"

3.2.3 가 가

1)

가

COD 가

2)

가
Table 3.4 3
가
가
pH 6.4
400mg/L
4000mg/L
가
가
가

Table 3.4 Employed in the nitrification process in this study

		Heavy metal concentration(mg/L)		
		Zn	Ni	Fe
Phase	1	10	5	5
	2	20	10	10
	3	40	20	20

3.2.4

1)

400mg/L

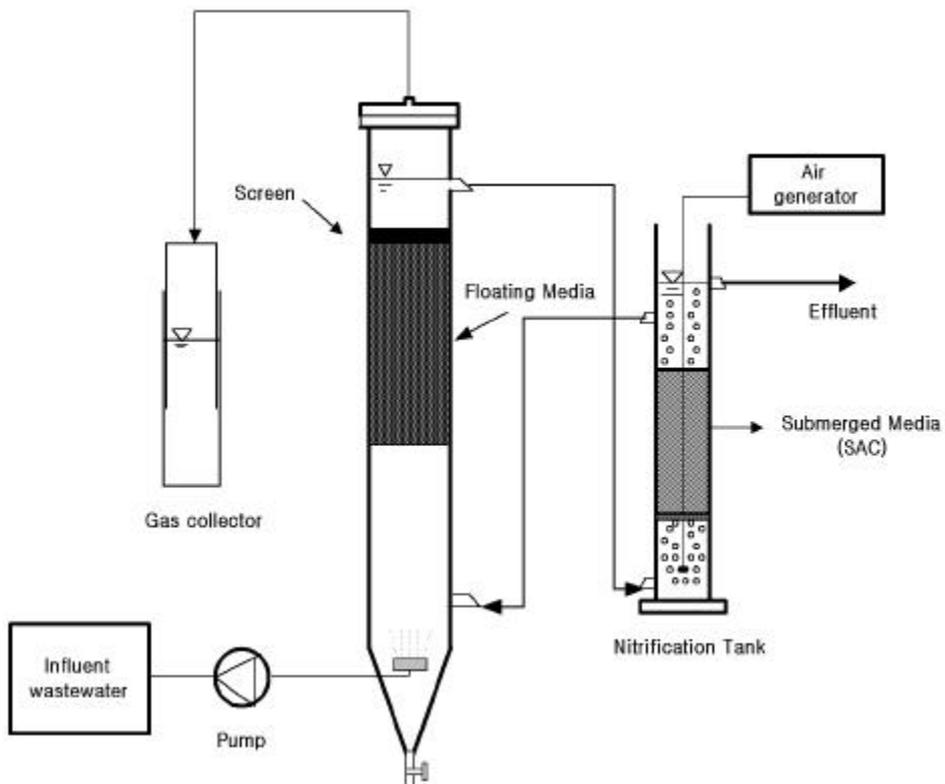


Fig. 3.2 Experimental apparatus for the denitrification system using Sulfate reducing bacteria, Sulfur denitrifier and Heterotrophic denitrifier.

1)

(Fig 3.2). 3가

가 가

(SO₄²⁻) 2000mg/L, (Alkalinity) 1000mg/L as CaCO₃ (by NaHCO₃), (NO₃-N) 50mg/L .

MPB

3.2.1

COD/Sulfate 0.5 .

200mg/L

3가

가

200mg/L

4000mg/L

가 .

3.2.1 .

A

가

가

3000mg/L

400mg/L

가 .

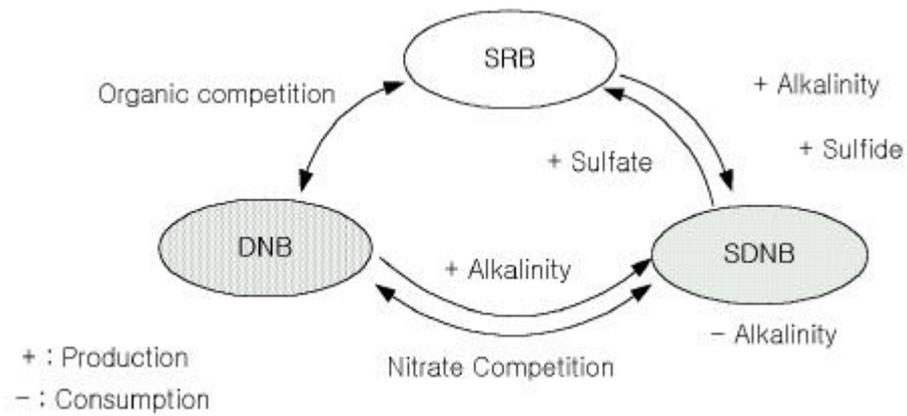


Fig. 3.3 Hypothetical diagram of relationships that occur among organism in denitrification reactor

3.3

3가

가

SRB

(DNB)

(SRB : Sulfate Reducing Bacteria)

(SDNB : Sulfur Denitrification

Bacteria)

가

. Fig. 3.3

SRB

SDNB

DNB

가

가

1) **SRB** :

1mg

1mg

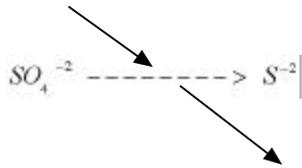
COD

0.4mg/L

. SRB

(19).

COD



Alkalinity

SO4 : COD : Alkalinity = - 1 : - 1 : +0.4

COD/sulfate ratio

SDNB :

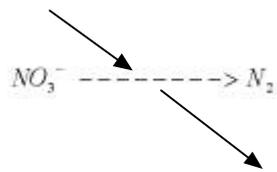
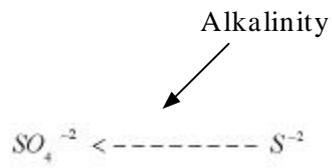
1mg

Sulfate 2.5mg

3.74mg

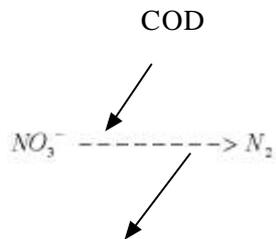
. SDNB

(4, 11).



NO₃⁻ : Sulfur(sulfate) : Alkalinity = - 1 : - 2.5(+2.5) : - 3.74

DNB :		1mg	COD
4.5mg	3.57mg		
(16).			



Alkalinity

NO₃⁻ : COD : Alkalinity = - 1 : - 4.5 : + 3.57

, COD,

Mass Balance for COD, Sulfate and Alkalinity to obtain each microbial role

COD : x	Sulfate : m
Sulfide : n	Alkalinity : y
Nitrate : z	1 for SRB
2 for SDNB	3 for DNB

$$x_1 + x_3 = \text{used COD} \text{-----} (3.1)$$

$$m_1 - m_2 = \text{removed sulfate(or } n_1 - n_2 = \text{effluent sulfide)-} (3.2)$$

$$y_1 - y_2 + y_3 = \text{removed alkalinity-} (3.3)$$

$$z_2 + z_3 = \text{removed nitrate-} (3.4)$$

3.4

2 3

0.45 μ m filter Sulfate,
 Ortho-P, Nitrate (Ion
 Chromatography, DX-500 equipped with CD-20)
 , TOC analyzer(Shmazu 5000)
 , Cr, Cu, Zn, Pb, Fe, Ni ICP-AES(ICP-IRIS)
 , 가 GC(equiped with TCD dector)
 Standard Methods
 (2).

4.1

COD/Sulfate

COD/sulfate

Sulfide

(19).

COD/sulfate

가

가

COD

Sulfate

COD/sulfate

2000ppm

Sulfate

COD/sulfate

4.1.1

가

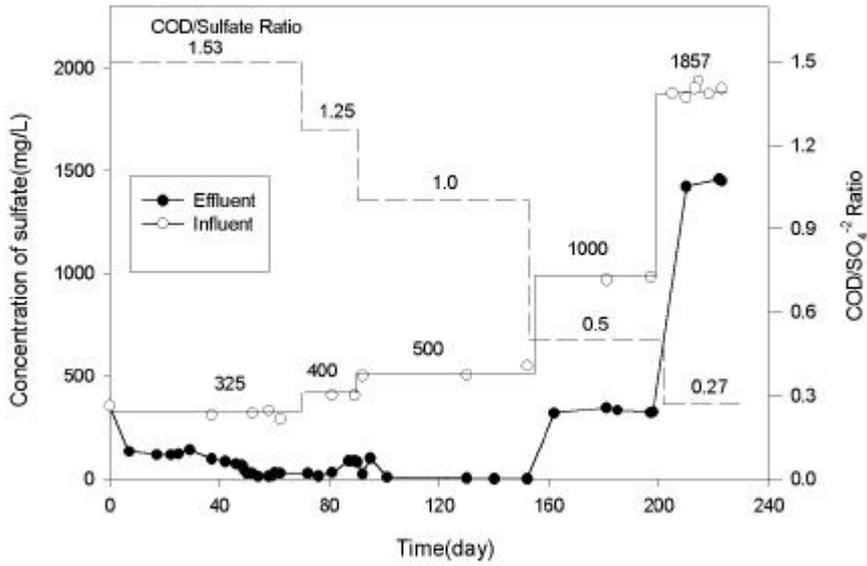


Fig. 4.1 Fate of sulfate and carbon source in the anaerobic sulfate-reducing reactor during the start-up period

가

500mg/L COD
 325mg/L COD/Sulfate 가 1.53
 HRT 1 . Fig.
 4.1 250
 10
 50% 가 , 50 COD가 150mg/L

(Table 4.1), 314mg SO₄2/L
 97% 가
 가 가 가
 가
 , 가 가
 0.4 COD/sulfate
 COD/sulfate 가 가
 , 1.7
 (19). COD/SO₄2 가 1.53 50

Table 4.1 Performance of the anaerobic sulfate-reducing reactor during the start-up period

COD/SO ₄ 2 ratio (mg/mg)	1.4- 1.6	0.98	0.51	0.26	1.0
Sulfate _{in} (mg/L)	325	500	1,000	1,857	1,000
Sulfate reduction(%)	96.9	98.0	67.0	28.0	80.0
Reduced sulfate(mg/L)	314	490	670	519	800
COD _{in} (mg/L)	500	500	500	500	1,000
COD _{eff} (mg/L)	150- 200	< 10	-	-	< 10

42mgAc/L
4.5mgAc/L
(19).

COD/sulfate
COD
1.53 COD/sulfate
COD 500mg/L
가
0.51- 1.03/d 0.11- 0.21/d , 1.37- 5.52/d
1.2- 2.0/d 가 (22).

(washout) 가
,
81
92 400mg SO₄2/L 500mg SO₄2/L
가 . 가
1 97%
, COD 10mg/L
160 200 1000mg SO₄2/L
1,857mg SO₄2/L 가 COD/sulfate 0.51 0.28
67%, 28% .

COD가

가 . , Fig.

4.1

COD/SO₄2

가 500mg/L

COD/sulfate

500mg COD/L

가

Table 4.2 Performance analysis of sulfate reducing reactor depending on the COD/SO42 ratio

COD/SO42 (mg/mg)	0.18	0.33	0.5	0.82	1.2
Influent SO42 (mg/L)	2,000	2,000	2,000	2,000	2,000
Sulfate reduction (%)	21.3	43	57.0	61	59
Amount of reduced sulfate(mg/L)	436	878	1,055	1,224	1,180
Influent COD(mg/L)	360	660	1,000	1,640	2,400
Effluent COD(mg/L)	-	-	< 10	400-600	1000-1,200

4.1.2

230

J

2,000mg/L

COD

COD/sulfate

가

Fig.

4.2 Table 4.2

COD/sulfate

COD/sulfate

0.18

COD

21.3% COD/sulfate 0.33 가
 COD , 43% 가
 , COD/sulfate
 (Fig. 4.2). 가 2,000mg/L COD/SO₄²⁻
 0.33

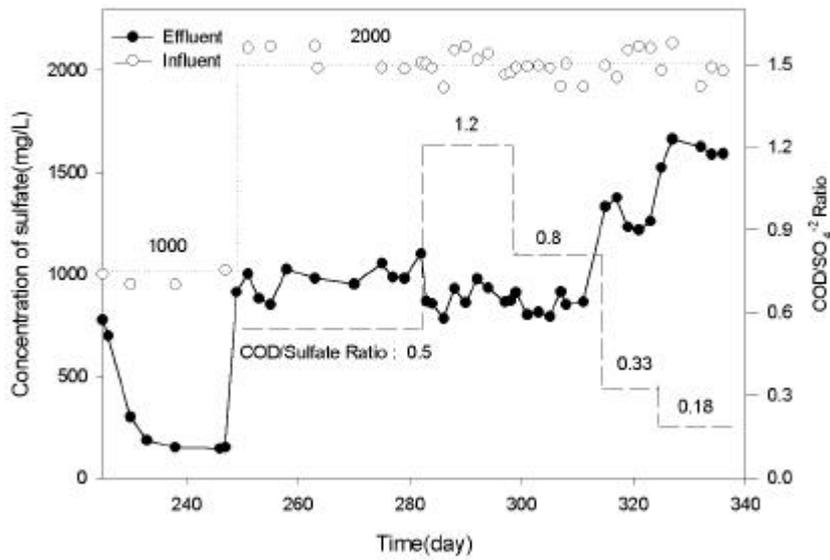


Fig. 4.2 Concentration of sulfate according to COD/sulfate ratio in the sulfate reducing reactor

COD/sulfate 가 0.5 57%
 COD가 10mg/L , COD/sulfate
 0.82 1.2 가 COD 600mg/L 1200mg/L
 가 61% 59% COD/sulfate 0.5

가 Table

4.2 COD 500mg/L COD/sulfate 0.51 0.98

67% 98% , 0.82

COD/sulfate COD가

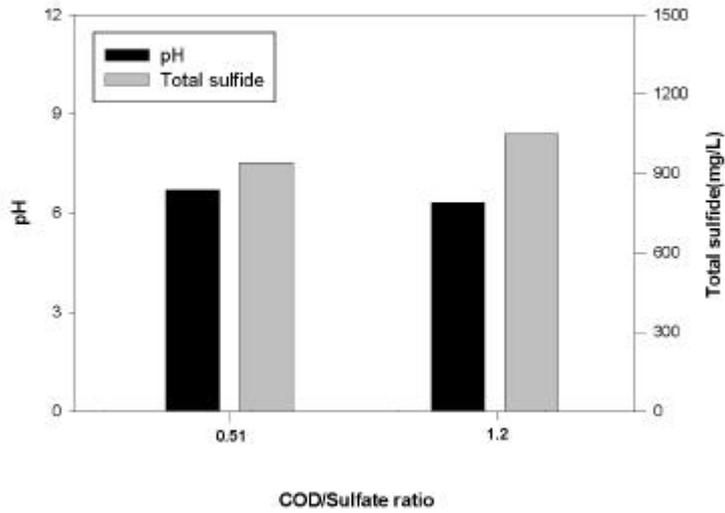


Fig. 4.3 pH and sulfide according to COD/SO4²⁻ ratio.

Fig. 4.3 COD/sulfate pH (Sulfide)

COD/sulfate 가 0.5 1.2

(Total sulfide) 940mg/L 1,050mg/L

(Total sulfide)

가 1,000mg/L

(19).

가 2,000mg/L

COD/SO4²⁻ 가 0.5

(S-2)

(HS-)

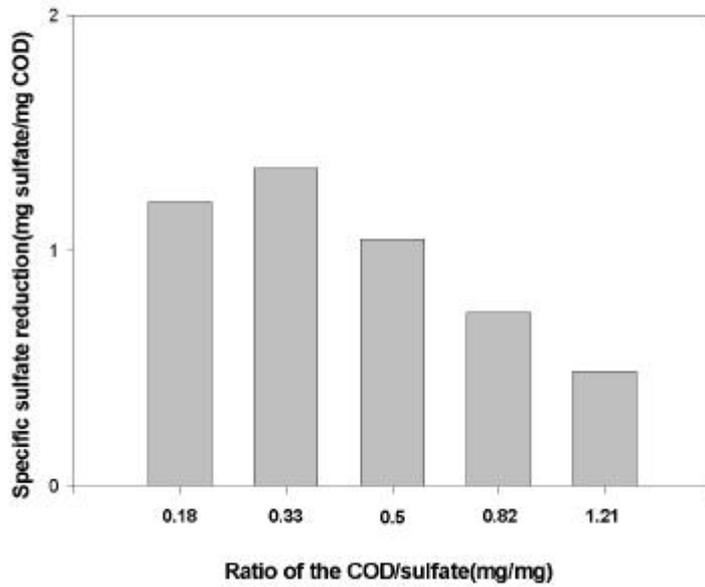


Fig. 4.4 Effectiveness of carbon source for according to COD/sulfate ratio

Fig. 4.4 COD/sulfate COD
(SO₄²⁻ mg/COD mg, Effectiveness of COD to reducing of sulfate)

COD
가 COD
COD/sulfate 가 가 COD/sulfate 0.33
Song (1998)

COD/sulfate 가 가 (39).

COD/sulfate
(scavenger) .

4.2

pH

가

(39). ,

가 . pH가 3.0

(Ni, Cr, Cu, Zn, Fe)

(Table 3.3), pH 5.4, 6.4 7.2

Fig. 4.5(a) pH

pH 5.4 23mg/L , 7.2

pH 가 가 . pH 5.4

104mg/L 가 가

pH 6.4 7.2 9.0mg/L 0.05mg/L

. pH 5.4 520mg/L pH 6.4

70mg/L pH 7.2

Fig.

4.5(b) (c)

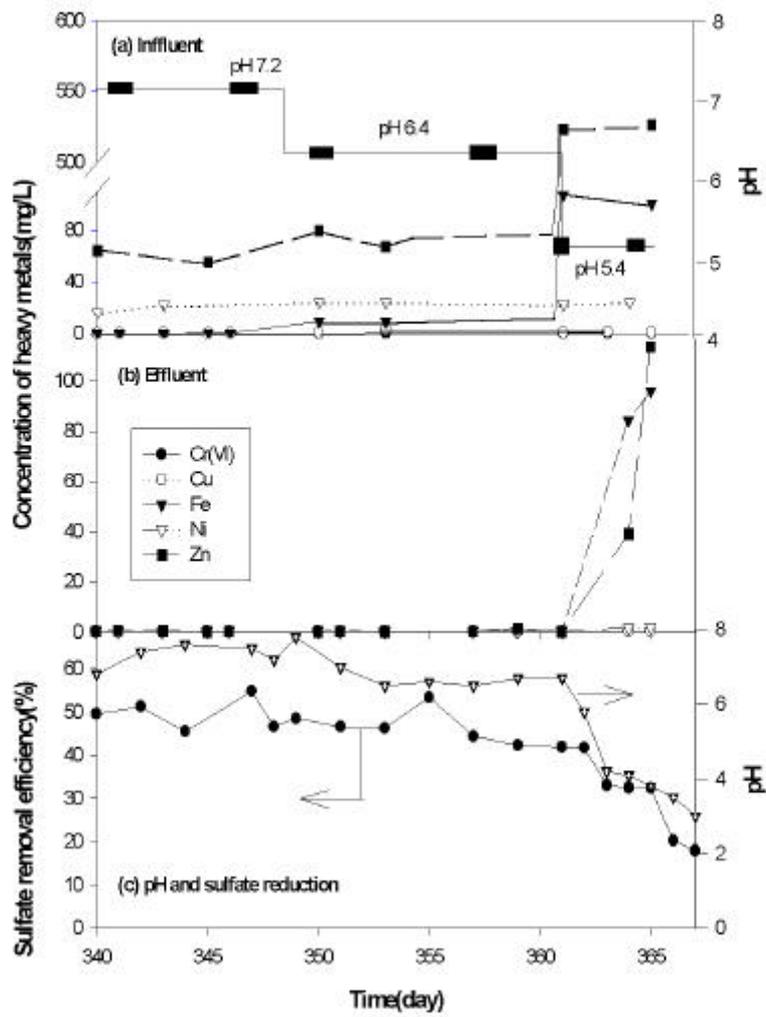
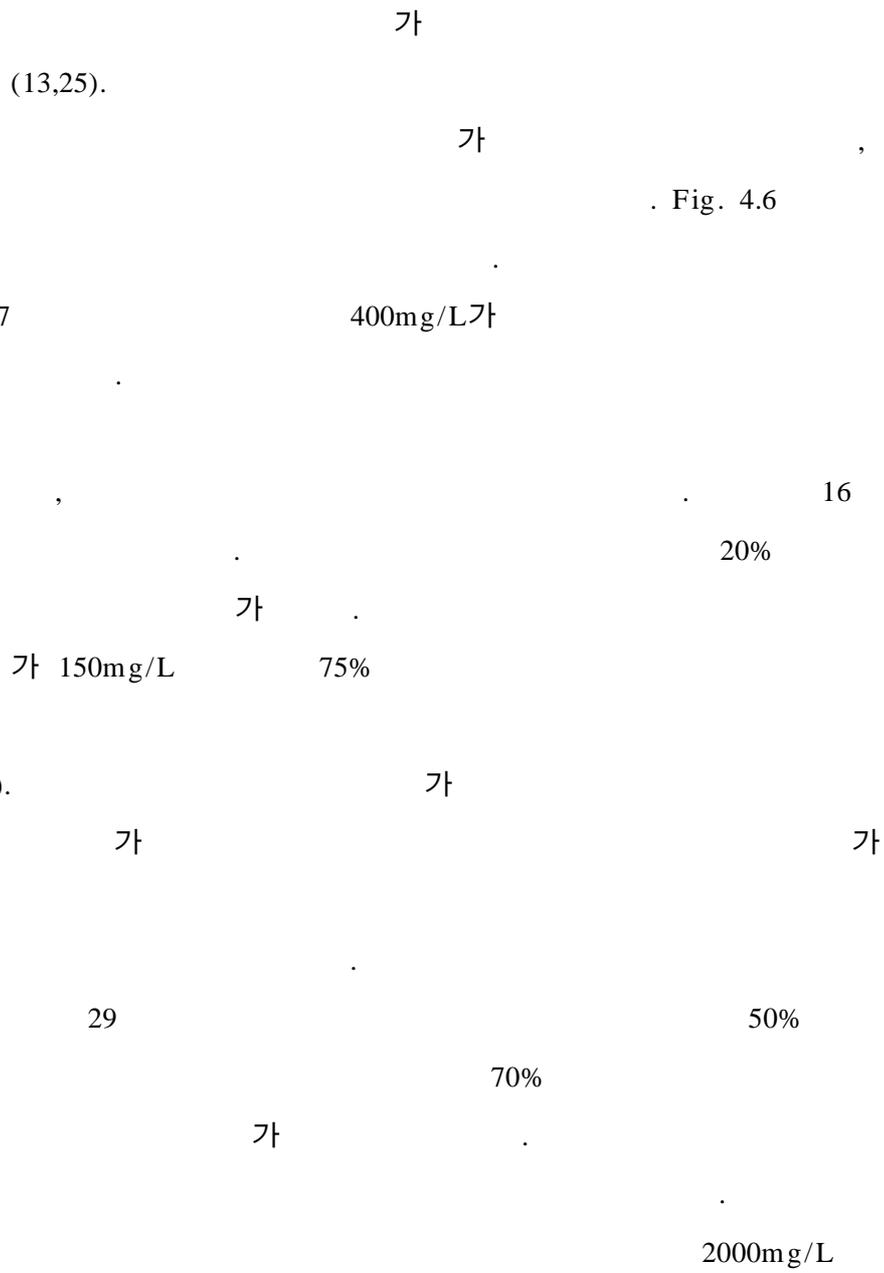


Fig. 4.5 Performance of heavy metal removal in the sulfate reducing reactor

4.3



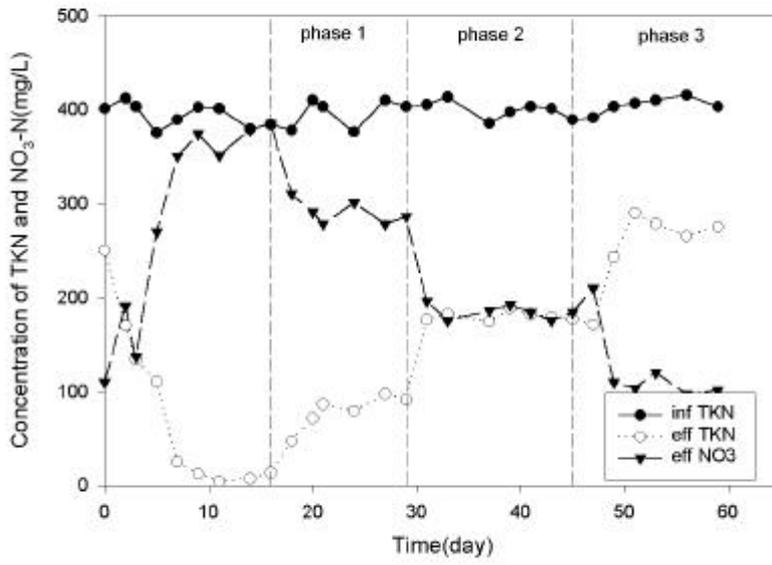


Fig 4.6 Time course of nitrate concentration in the nitrification reactor

4.4

가

Fig. 4.7

B

41 50mg/L

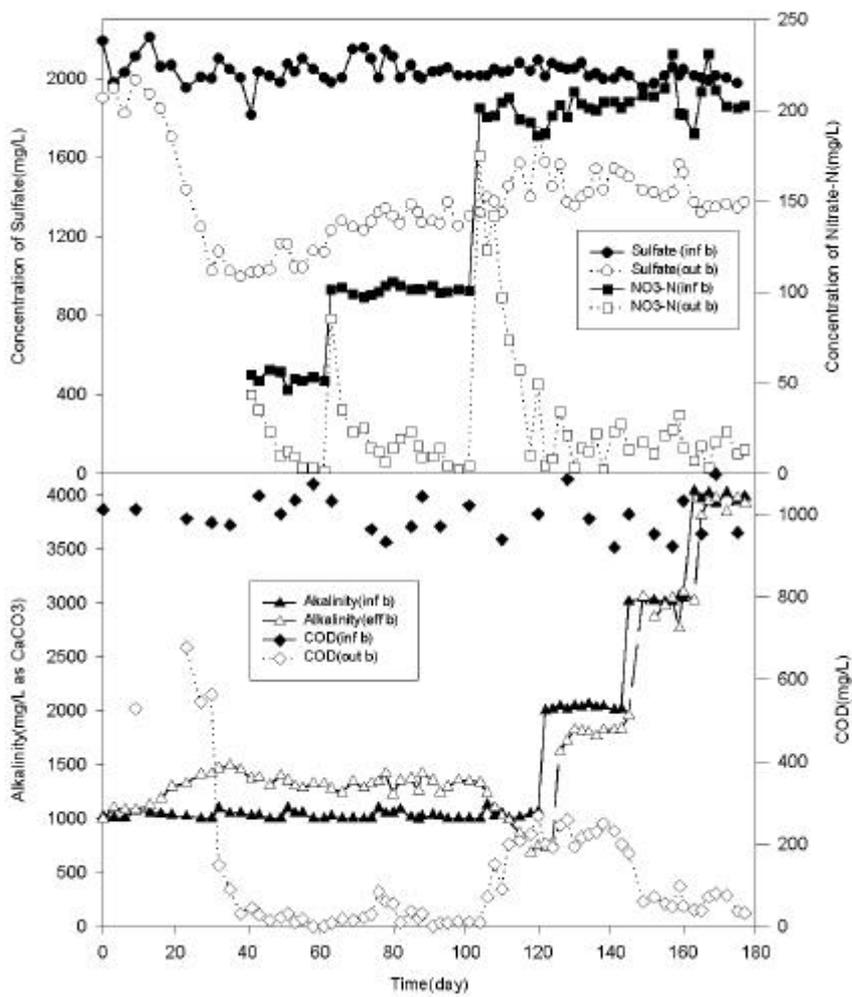


Fig.4.7 Behaviors of denitrification process by sulfur denitrifier and heterotrophic denitrifier according to the increase of nitrate(Reactor B)

10

가 (SRB)

, SRB

DNB

,
SDNB

가

COD/N 가 10 DNB

Ammonification

(30).

Ammonification

63

100mg/L

14

가 10mg/L

2

0 30%

SRB

COD

가

SRB

200 300mg/L COD DNB가

45 65mg/L가 DNB 가

1220 1350mg/L

100mg/L

DNB가 40 65%, SDNB가

가 102 200mg/L

40 50%

COD가 190 250mg/L

15

가

DNB가 50 70mg/L 가

COD DNB가

COD
 COD
 200mg/L COD 가
 B 가
 가
 2,000mg/L 1,000mg/L
 3,000mg/L 145
 COD 50 100mg/L가
 80 90mg/L DNB
 balance
 4000mg/L COD
 SRB 65% 가
 SDNB가 130 140mg/L
 4000mg/L
 가
 2500 3000mg/L 가

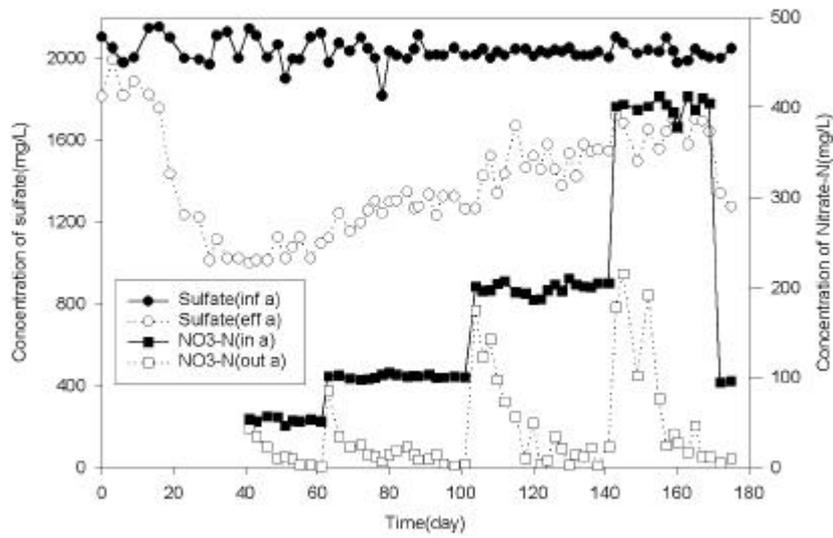


Fig. 4.8. Behaviors of denitrification process by sulfur denitrifier and heterotrophic denitrifier according to the increase of nitrate(Reactor A)

Fig 4.8 A . 143
 400mg/L 3000mg/L COD 1000mg/L
 . 60 70% 가
 COD .
 200mg/L COD DNB
 가 . 12
 가 30mg/L 가
 .
 DNB 130 160mg/L SDNB 가
 가 .
 3000mg/L COD 1000mg/L 400mg/L

Sulfur cycle 가 가 가 . SRB
60~70% DNB 가

가 SRB .
가

1.

500mg COD/L

325mg SO₄²/L

가

가 .

2.

2,000mg/L

COD/SO₄²

0.33

가

,

COD/SO₄² 0.5

.

3.

COD/SO₄²

0.33

가 .

4.

COD/SO₄²

,

COD/SO₄²

.

,

가 ,

5.

pH 6.4

pH 5.4

pH

pH 5.4

6.

가

7.

3000mg/L, COD

1000mg/L

SRB, SDNB, DNB

가

1. , 3가

2. HRT 가

3. Lab scale

가 .

4. 가 .

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